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(Supersedes ADA213392)

GRANT NO: DAMD17-89-Z-9021

TITLE: DISCOVERY AND DEVELOPMENT OF THERAPEUTIC DRUGS AGAINST

LETHAL HUMAN RNA-VIRUSES: A MULTIDISCIPLINARY ASSAULT

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REPORT DATE: February 20, 1990

TYPE OF REPORT: Midterm Report



PREPARED FOR: U.S. ARMY MEDICAL RESEARCH AND DEVELOPMENT COMMAND

Fort Detrick, Frederick, Maryland 21701-5012

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REPORT DOCUMENTATION PAGE							Form Approved OMB No. 0704-0188			
Unclas				16. RESTRICTIVE MARKINGS						
2a. SECURITY	CLASSIFICATIO	N AUTHORITY		3 DISTRIBUTION / AVAILABILITY OF REPORT						
2b. DECLASSI	FICATION / DOV	WNGRADING SCHEDU	LE		for public : ion unlimite		± <b>;</b>			
4. PERFORMING ORGANIZATION REPORT NUMBER(S)				5. MONITORING	ORGANIZATION RI	PORT NU	M8ER(S)			
		ORGANIZATION	6b. OFFICE SYMBOL (If applicable)	7a. NAME OF M	ONITORING ORGA	NIZATION				
	Research			7						
6c. ADDRESS (City, State, and ZIP Code)			76. ADDRESS (Cit	ty, State, and ZIP C	(ode)					
Arizona State University Tempe, Arizona										
8a. NAME OF	FUNDING / SPO	ONSORING Army Medical	8b. OFFICE SYMBOL (If applicable)	9. PROCUREMEN	T INSTRUMENT IDE	NTIFICATI	ON NUMBER			
		ment Command	(II applicable)	Contract N	lo. DAMD17-89	9-Z-902	:1			
8c. ADDRESS (	City, State, and	d ZIP Code)	l	10 SOURCE OF F	UNDING NUMBER	S				
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Frederick	, Marylan	d 21701-5012		ELEMENT NO. 62787A	NO. 3M1- 62787A871	NO.	ACCESSION NO.			
11. TITLE (Incl	ude Security C	lassification)		027078	02/0/A0/1		374			
	<del>-</del>		THERAPEUTIC DRUG	S AGAINST LE	THAL HUMAN I	RNA-VIF	RUSES:			
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12. PERSONAL	• •									
13a. TYPE OF	R. Pettit	13b. TIME CO	OVERED	14 DATE OF BERO	RT (Year Month I	Dayl 15	PAGE COUNT			
Midterm			5/89 <b>to</b> 2/5/90	14 DATE OF REPORT (Year, Month, Day) 15. PAGE COUNT 1990 February 20 25						
16. SUPPLEME Superse		MON ADA213392.								
17	COSATI	CODES	18. SUBJECT TERMS (	Continue on revers	e if necessary and	identify b	y block number)			
FIELD	GROUP	SUB-GROUP	RA I; Antivir							
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The grant funds have been sharply focused on discovery of new antiviral drugs. We have been pursuing the pancratistatin family of antiviral leads as a top priority. The research results here have been very encouraging and pancratistatin, isonarciclasine, cis-dihydronarciclasine as well as trans-dihydronarciclasine have proved to be quite promising. Meanwhile over 1700 naturally occurring specimens are now undergoing preliminary antiviral evaluation at USAMRIID. The resulting leads will also be pursued as rapidly as financial resources permit. The most exciting overall result has been the discovery in USAMRIID's laboratories that pancratistatin will cure the in vivo experimental version of Japanese Encephalitis. In addition, of the 1770 samples submitted during this period for pre-screen testing, we have received data for 875. Of these 150 samples displayed possible activity against either PT or YF or both for a total of 173 positive results.  21 ABSTRACT SECURITY CLASSIFICATION    UNCLASSIFIED/UNLIMITED   SAME AS RPT   DIIC USERS   DIIC USERS										
22a. NAME OF	RESPONSIBLE	INDIVIDUAL			Include Area Code)					
Mrs. Vi	rginia M.	Miller		301/663-7			GRD-RMI-S			

#### **FOREWORD**

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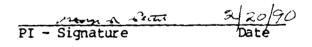
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For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

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#### INTRODUCTION

To summarize, a long-term USAMRIID research program directed at the isolation and structural elucidation of new and potentially useful antiviral drugs from marine animals and plants is in progress. The financial support provided by the USAMRIID program will continue to be used to isolate and characterize new antiviral chemotherapeutic drugs from confirmed active extracts of marine invertebrates and vertebrates as well as marine and terrestrial plants including fungi, algae and other microorganisms. The research is sharply directed at marine animal and plant species yielding extracts with an outstanding level of antiviral activity in the USAMRIID's programs (RNA viruses).

#### BODY

The overall results and status to date are as follows. The grant funds are being used for discovery of new antiviral drugs. We have been pursuing the pancratistatin family of antiviral leads as a top priority. The research results here have been very encouraging and pancratistatin, isonarciclasine, cis-dihydronarciclasine as well as trans-dihydronarciclasine have proved to be quite promising. Meanwhile over 1700 naturally occurring specimens are now undergoing preliminary antiviral evaluation at USAMRIID. The most exciting overall result has been the discovery in USAMRIID's laboratories that pancratistatin will cure the in vivo experimental version of Japanese Encephalitis. In addition, of the 1770 samples submitted during this period for pre-screen testing, we have received data for 875. Of these 150 samples displayed possible activity against either PT or YF or both for a total of 173 positive results.

To follow is a detailed report of the latest antiviral evaluation results as Part A. In part B appears experimental results corresponding to our synthetic transformations aimed at uncovering important antiviral derivatives of pancratistatin and developing a workable synthetic approach to pancratistatin based on narciclasine. Meanwhile we have completed reisolation of pancratistatin from 1/2 ton of Pancratium littorale collected in the Republic of Seychelles. Enough pancratistatin is now available for the next series of in vivo experiments. In short progress continues to be excellent and we have a magnificent number of new antiviral leads to pursue.

Also an outline summary of fractions obtained from the Papua New Guinea sponge B723344 and the Philippines marine sponge B721160 is attached.

A major effort has been devoted to scaling-up the isolation of narciclasine from a <u>Narcissus</u> species. We will shortly have 10 grams of narciclasine ready for chemical conversions and we will be concentrating on increasingly larger scale-up operations using the <u>Narcissus</u> with the objective of obtaining about 100+ grams of narciclasine.

In summary we are making excellent progress and this has been made possible by the most necessary USAMRIID support.

# $\underline{P} \underline{A} \underline{R} \underline{T} \underline{A}$

ASU NUM	AREA	ТУРЕ	VIRUS	DATA	SCREEN	DATE RECI	DRUG NUM	AVS NUM
B23-110			YF	22.03	Pre	11/14/89		4855
B23-110			PT	12.27		11/14/89		4855
M-2927	Philippines.79	Echinodermata	PT	12.35		09/28/89	B721092	1312
M-2928	Philippines.79		PT	2.21		09/28/89	B721095	1315
M-3070	Philippines.79		PT	2.23		11/14/89	B721511	1801
M-3077	Philippines.79		PT	1.19		09/28/89	B721532	1818
M-3093	Philippines.79		PT	13,49		09/28/89	B721568	1850
M-3093	Philippines.79		PT	34.08		11/14/89	11	1850
M-3093	Philippines.79	Porifera	YF	22.24		09/28/89	11	1850
M-3312	Australia.78	Coelenterata	PT	1.90		11/14/89	B721743	
M-3314	Australia,78	Coelenterata	PT	2.18	Pre	11/14/89	B721749	6584
M-3328	Australia.78	Chordata/Tunicata	PT	1.67	Pre	10/31/89	B721787	6585
M-3136	Palau.79	Mollusca	PT	1.33	Pre	10/31/89	B721818	6586
M-3137	Palau.79	Porifera	PT	5.16	Pre	10/31/89	B721823	6248
M-3137	Palau.79	Porifera	PT	7.29	Pre	11/20/89	/1	6248
M-3137	Palau.79	Porifera	YF	6.51	Pre	10/31/89	14	6248
M-3137	Palau.79	Porifera	YF	1.74	Pre	01/02/90	B	6248
M-3138	Palau.79	Porifera	PT	1.82		10/31/89	B721826	6587
M-3142	Palau.79	Echinodermata	PT	3.73		10/31/89	B721838	6588
M-3156	Palau.79	Porifera	PT	13.99		10/31/89	B721880	6589
M-3160	Palau.79	Chordata/Pisces	PT	1.02		10/31/89	B721892	6590
M-3163	Palau.79	Chordata/Pisces	YF	10.52		01/02/90	B721899	6591
M-3163	Palau.79	Chordata/Pisces	PT	5.84		01/02/90	11	6591
M-3165	Palau.79	Chordata/Pisces	PT	5.56		01/02/90	B721905	6592
M-3166	Palau.79	Chordata/Pisces	PT	3.29		11/09/89	B721908	6593
M-3166	Palau.79	Chordata/Pisces	PT	3.56		11/09/89	B721910	6594
M-3169	Palau.79	Chordata/Pisces	PT	7.41		11/09/89	B721917	6595
M-3171	Palau.79	Chordata/Pisces	PT	2.16		11/09/89	B721925	6596
M-3181	Palau.79	Chordata/Pisces	PT	2.69		01/02/90	B721953	6597
M-3181	Palau. 79	Chordata/Pisces	YF	3.13		01/02/90	)  	6597
M-3182	Palau.79	Chordata/Pisces	PT	1.63		11/20/89	B721958	6598
M-3189	Palau.79	Chordata/Pisces	PT	3.35 1.32		01/02/90	B721979 B722006	6599
M-3198 M-3212	Palau.79 Palau.79	Chordata/Pisces Porifera	PT PT	8.13		11/20/89	B722048	6600 6601
M-3212	Palau.79	Porifera	PT	3.10		01/02/90 01/02/90	B722048	6602
M-3214	Palau.79		PT	27.94		01/02/90	B722054	6603
M-3222	Palau.79		PT	4.96		01/09/90	B722076	6604
M-3222	Palau.79		PT	26.75		01/09/90	B722077	6605
M-3222	Palau.79	Porifera	PT	3.49		01/09/90	B722078	6606
M-3223	Palau.79		PT	4.05		01/09/90	B722080	6607
M-3223	Palau.79		PT	14.03		01/09/90	B722081	6608
M-3225	Palau.79		PT	1.97		01/02/90	B722087	6609
M-3225	Palau.79		YF	1.05		01/02/90	rt.	6609
M-3226	Palau.79		PT	1.19		01/02/90	B722089	6610
M-3227	Palau.79		PT	11.17		01/02/90	B722091	6611
M-3228	Palau.79		PT	5,28		01/09/90	B722094	6612
M-3233	Palau. 79		YF	1.71		01/09/90	B722109	6613
M-3233	Palau, 79		YF	3.85		01/09/90	B722111	6614
M-3243			PT	1.69		01/09/90	B722141	6615
M-3250	Palau.79		PT	1.23		01/09/90	B722162	6616
M-3251	Palau.79		PT	1.46		01/09/90	B722165	6617

N-3251   Palau.79   Porifera   YF   2.90   Pra   01/09/90   7618   6618   N-3257   Palau.79   Porifera   YF   1.63   Pra   01/09/90   B722168   6618   N-3257   Palau.79   Porifera   YF   1.31   Pra   01/08/90   B722162   6629   N-3257   Palau.79   Porifera   PT   2.68   Pra   01/16/90   7620   6620   N-3257   Palau.79   Porifera   PT   7.13   Pra   01/16/90   7620   6620   N-3257   Palau.79   Porifera   PT   7.13   Pra   01/16/90   7620   N-3257   Palau.79   Porifera   PT   7.48   Pra   01/16/90   7621   6621   N-3271   Palau.79   Porifera   PT   13.02   Pra   01/08/90   B722222   6622   N-3271   Palau.79   Porifera   PT   14.07   Pra   01/08/90   B722222   6624   N-3273   Palau.79   Porifera   PT   14.07   Pra   01/08/90   B722224   6623   N-3273   Palau.79   Porifera   PT   2.41   Pra   01/08/90   B722224   6624   N-3273   Palau.79   Porifera   PT   2.41   Pra   01/08/90   B722239   6676   N-3276   Palau.79   Porifera   PT   3.12   Pra   01/08/90   B722230   6625   N-3277   Palau.79   Porifera   PT   3.12   Pra   01/08/90   B722230   6626   N-3277   Palau.79   Porifera   PT   3.12   Pra   01/08/90   B722246   6628   N-3279   Palau.79   Porifera   PT   2.62   Pra   01/08/90   B722246   6628   N-3279   Palau.79   Porifera   PT   2.62   Pra   01/08/90   B722246   6628   N-3279   Palau.79   Porifera   PT   1.69   Pra   01/08/90   Pra   6627   N-3277   Palau.79   Porifera   PT   1.69   Pra   01/08/90   Pra   6628   N-3279   Palau.79   Porifera   PT   1.69   Pra   01/08/90   Pra   6628   N-3279   Palau.79   Porifera   PT   1.69   Pra   01/08/90   Pra   6628   N-3279   Palau.79   Porifera   PT   1.69   Pra   01/08/90   Pra   6629   N-3279   Palau.79   Porifera   PT   1.69   Pra   01/08/90   Pra   6629   N-3279   Palau.79   Porifera   PT   1.69   Pra   01/08/90   Pra   6629   N-3279   Palau.79   Porifera   PT   1.69   Pra   01/08/90   Pra   6629   N-3279   Pra   01/08/90   Pra   6629   N-3279   Pra   01/08/90   Pra   6629   N-3279   Pra   01/08/90   Pra   6629   Pra   6629   Pra   6629   Pra   6629   Pra   6629   P	ASU NUM	AREA	TYPE	VIRUS	DATA	SCREEN	DATE RECD	DRUG NUM	AVS NUM
M-3251 Palau.79 Porifera YF 1.31 Pre 01/09/90 B722181 6619 M-3257 Palau.79 Porifera YF 1.31 Pre 01/09/90 B722182 6620 M-3257 Palau.79 Porifera YF 44.17 Pre 01/16/90						<b>n</b>	01 (00 (00		6617
N-1257 Palau.79 Porifera PT 2.68 Pre 01/08/90 B722182 6620 N-3257 Palau.79 Porifera PT 7.13 Pre 01/08/90 B722182 6620 N-3257 Palau.79 Porifera PT 7.13 Pre 01/08/90 B722183 6620 N-3257 Palau.79 Porifera PT 7.13 Pre 01/08/90 B722183 6621 N-3257 Palau.79 Porifera PT 7.13 Pre 01/08/90 B722222 6622 N-3271 Palau.79 Porifera PT 13.02 Pre 01/08/90 B722222 6622 N-3271 Palau.79 Porifera PT 14.07 Pre 01/08/90 B722223 6624 N-3273 Palau.79 Porifera PT 4.07 Pre 01/08/90 B722223 6625 N-3273 Palau.79 Porifera PT 4.07 Pre 01/08/90 B722223 6626 N-3273 Palau.79 Porifera PT 4.07 Pre 01/08/90 B722223 6626 N-3276 Palau.79 Porifera PT 4.07 Pre 01/08/90 B722239 6626 N-3277 Palau.79 Porifera PT 4.07 Pre 01/08/90 B722239 6626 N-3277 Palau.79 Porifera PT 5.60 Pre 01/08/90 B722239 6626 N-3277 Palau.79 Porifera PT 1.12 Pre 01/08/90 B722239 6626 N-3277 Palau.79 Porifera PT 2.26 Pre 01/08/90 B722244 6627 N-3277 Palau.79 Porifera PT 2.26 Pre 01/08/90 B722246 6628 N-3279 Palau.79 Porifera PT 2.26 Pre 01/08/90 B722246 6628 N-3279 Palau.79 Porifera PT 1.47 Pre 01/08/90 B722446 6628 N-3279 Palau.79 Porifera PT 1.49 Pre 01/08/90 B722446 6628 N-3279 Palau.79 Porifera PT 1.49 Pre 01/08/90 B722446 6628 N-3279 Palau.79 Porifera PT 1.47 Pre 01/08/90 B722446 6628 N-3279 Palau.79 Porifera PT 1.49 Pre 01/08/90 B722446 6628 N-3279 Palau.79 Porifera PT 1.49 Pre 01/08/90 B722446 6628 N-3532 Wewak.85 Porifera PT 1.50 Pre 09/28/89 B724332 6631 N-5533 Wewak.85 Porifera PT 1.50 Pre 09/28/89 B724336 6631 N-5535 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724336 6634 N-5556 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724336 6634 N-5557 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724336 6637 N-5557 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724336 6637 N-5556 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724436 6637 N-5556 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724436 6637 N-5557 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724436 6637 N-5558 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724436 6637 N-5558 Wewak.85 Porifera PT 1.51 Pre 09/28/89 B724436 6637 N-5558 Wewak.85 Porifera PT 1.52 Pre 0									
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M-5603 Wewak.85 Porifera PT 6.31 Pre 09/28/89 B724455 6655			Porifera						
			Porifera				•		
M-5605 Wewak.85 Portiera		Wewak.85	Porifera	PT					
M-5606 Wewak.85 Porifera PT 10.48 Pre 09/28/89 B/24436 0030			Porifera				•		
M-5607 Wewak.85 Porifera PT 4.55 Pre 09/28/89 B/2445/ 005/			Porifera				•		
M-5608 Wewak.85 Porifera PT 7.84 Pre 09/28/89 B724458 6658			Porifera	PT	7.84	rre	09/28/89	D/24438	טנטט

ASU NUM	AREA	TYPE	VIRUS	DATA	SCREEN	DATE	RECD	DRUG N	UM	AVS	NUM
M-5616	Wewak.85	Porifera	PT	6.11	Pre	09/28	/89	B72446	6	6659	)
M-5616	Wewak.85	Porifera	YF	4.77		09/28		11		6659	)
M-5618	Wewak.85	Porifera	ΥF	3.33	Pre	09/28	/89	B72446	8	6660	
M-5658	Wewak.85	Porifera	PT	3.64	Pre	11/14	/89	B72450	8	6661	
M-5659	Wewak.85	Echinodermata	PT	1.28	Pre	11/14	/89	B72450	9	6662	
M-5662	Wewak.85	Porifera	PT	5.77	Pre	11/14	/89	B72451	2	6663	
M-5667	Wewak.85	Porifera	PT	1.17	Pre	11/14	/89	B72451	7	6664	
M-5669	Wewak.85	Porifera	PT	6.58	Pre	11/14	/89	B72451	9	6665	
M-5671	Wewak.85	Porifera	PT	11.38	Pre	11/14	/89	B72452	1	6666	
M-5675	Wewak.85	Porifera	PT	1.18	Pre	11/14	/89	В72452	5	6667	
M-5676	Wewak.85	Porifera	PT	28.78	Pre	11/14	/89	B72452	6	6668	
M-5677	Wewak.85	Porifera	PT	7.75	Pre	11/14	/89	B72452	7	6669	
M-5680	Wewak.85	Porifera	YF	1.84	Pre	11/14	/89	B72453	0	6670	
M-5680	Wewak.85	Porifera	PT	1.26	Pre	11/14	/89	11		6670	)
M-5685	Wewak.85	Porifera	PT	1.44		11/14	/89	B72453	5	6671	•
M-5694	Wewak, 85	Porifera	PT	1.47		11/14	/89	B72454	4	6672	
M-5736	Wewak.85	Porifera	PT	4.57	Pre	09/28	/89	B72458	6	6677	
M-5740	Wewak.85	Coelenterata	PT	2.71		09/28	/89	B72459	0	6678	;
M-5742	Wewak.85	Porifera	PT	3.34		09/28	-	B72459	2	6679	)
M-5742	Wewak.85	Porifera	YF	4.94		09/28	-	11		6679	)
M-5742	Wewak.85	Porifera	PT	3.44		09/28	-	B72459	6	6680	)
M-5757	Wewak.85	Porifera	PT	3.76		09/28	-	B72460	7	6681	
M-5760	Wewak.85	Porifera	YF	5.82		09/28		B72461	0	6682	
M-5768	Wewak.85	Porifera	YF	6.69		09/28		B72461	8	6683	
M-5777	Wewak.85	Porifera	PT	2.06	Pre	09/28	/89	B72462	7	6684	
M-5783	Wewak.85	Echinodermata	PT	2.32	Pre	09/28	/89	B72463	3	6685	
M-5792	Wewak.85	Porifera	PT	1.23	Pre	09/28	/89	B72464	2	6686	
M-5794	Wewak.85	Porifera	PT	2.77	Pre	09/28	/89	B72464		6687	
M-5802	Wewak.85	Echinodermata	PT	3.28	Pre	09/28	/89	B72465		6688	
M-5804	Wewak.85	Porifera	PT	1.21	Pre	09/28	/89	B72465		6689	
M-5808	Wewak.85	Porifera	PT	1.52	Pre	09/28	/89	B72465		6690	
M-5812	Wewak.85	Echinodermata	PT	1.38	Pre	09/28		B72466		6691	
M-5815	Wewak.85	Porifera	PT	2.36		09/28		B72466		6692	
M-5818	Wewak.85	Porifera	PT	1.57	Pre	10/31		B72466		6693	
	Wewak.85	Porifera	PT	4.44	Pre	10/31		B72467		6250	
M-5848	Wewak.85	Porifera	PT	5.38	Pre	10/31		В72469		6251	
M-5849	Wewak.85	Porifera	PT	11.19	Pre	10/31		B72469		6252	
M-5852	Wewak.85	Porifera	YF	5.39		10/31	-	B72470		6253	
M-5861	Wewak.85	Porifera	PT	4.77		10/31	-	B72471		6254	
M-5863	Wewak.85	Porifera	PT	2.90		10/31		B72471		6255	
M-5865	Wewak.85	Porifera	PT	1.16		10/31	•	B72471		6256	
M-5868	Wewak.85	Porifera	PT	2.97		10/31		B72471		6257	
M-5869	Wewak.85	Porifera	PT	1.46		10/31		B72472		6258	
M-5871	Wewak.85	Porifera	PT	6.12		10/31	•	B72472		6259	
M-5873	Wewak.85	Porifera	PT	2.56		10/31		B72472		6694	
M-5877	Wewak.85	Porifera	PT		Pre	10/31		B72472		6695	
M-5878	Wewak.85	Porifera	PT	2.04		10/31		B72472		6696	
M-5881	Wewak.85	Porifera	PT		Pre	10/31		B72473		6697	
M-5889	Wewak.85	Porifera	PT	20.27		10/31	-	B72474		6261	
M-5912	Wewak.85	Porifera	PT	4.95		10/31		B72476		6262	
M-5914	Wewak.85	Porifera	PT	6.41	Pre	10/31	./89	B72476	94	6263	,

ASU NUM	AREA	TYPE	VIRUS	DATA	SCREEN	DATE	RECD	DRUG	NUM	AVS	NUM
M-5919	Wewak.85	Chord./Elasmobra.	PT	1.59	Pre	10/3	•	B7247		6264	
M-5922	Wewak.85	Chord./Elasmobra.	YF	1.26	Pre	10/3	•	B7247		6698	
M-4894	Maldives.86	Porifera	PT	5.55	Pre	10/3	L/89	B7247	781	626	
M-4896	Maldives.86	Porifera	PT	5.69	Pre	10/3	L/89	B7247	783	6699	
M-4898	Maldives.86	Porifera	PT	2.21	Pre	10/3	L/89	B7247	785	6700	
M-4910	Maldives.86	Porifera	PT	2.07	Pre	10/33	L/89	B7247		6266	
M-4925	Maldives.86	Coelenterata	YF	1.69	Pre	10/3	L/89	B7248		670	
M-4933	Maldives.86	Porifera	PT	6.48	Pre	10/33	L/89	B7248		6261	
M-4938	Maldives.86	Porifera	PT	32.91	Pre	10/3	L/89	B7248	325	6269	
M-4945	Maldives.86	Porifera	PT	1.50	Pre	10/3	L/89	B7248	332	670	
M-4957	Maldives.86	Porifera	YF	2.18	Pre	10/31	L/89	B7248	344	6270	
M-4966	Maldives.86	Porifera	YF	2.09	Pre	10/31	L/89	B7248	352	6272	
M-4969	Maldives.86	Porifera	PT	3.13	Pre	10/3	L/89	B7248	355	670	3
M-4974	Maldives.86	Porifera	PT	3.12	Pre	10/31	L/89	B7248	360	6274	4
M-4977	Maldives.86	Porifera	YF	1.81	Pre	10/31	1/89	B7248	363	627	5
M-4977	Maldives.86	Porifera	PT	2.69	Pre	10/33	1/89	"		627	5
M-4980	Maldives.86	Porifera	PT	1.12	Pre	10/31	1/89	B7248	366	6704	<b>,</b>
M-5001	Maldives.86	Porifera	PT	1.57		10/31	/89	B7248	885	6709	5
M-5002	Maldives.86	Porifera	PT	7.84		10/31	L/89	B7248	886	6277	7
M-5002 M-5013	Maldives.86	Porifera	YF	8.65		10/31	•	B7248	198	6706	<b>်</b>

## SEHI-SYNTHETIC APPROACHES TO PANCRATISTATIN

#### Narciclasine-3,4-acetonide

To a solution of narciclasine (1.0 g, 3.25 mmol) in dimethylformamide (5 mL) and dimethoxypropane (5 mL) was added p-toluene sulfonic acid (100 mg). The solution was stirred at room temperature overnight. Acetonide precipitated out of solution. Pyridine (1 mL) and water (50 mL) was added and the mixture was stirred at room temperature for 30 minutes. The precipitate was collected by filtration, washed with water and dried at 64°C over  $P_2O_5$  under high vacuum to give as an amorphous powder, narciclasine-3,4-acetonide (1.05 g, 92.9%), mp. 275-7°, IR (NaCl)  $\nu_{\rm max}$  3500, 3150, 1637, 1625, 1596, 1464, 1437, 1337, 1201, 1079, 1038, 1019 cm<sup>-1</sup>, <sup>1</sup>HNMR & (CDCl<sub>3</sub>) 1.39 (s, 3H, CH<sub>3</sub>), 1.53 (s, 3H, CH<sub>3</sub>), 2.47 (d, J = 4.1 Hz, 1H, OH), 4.10-4.13 (m, 3H, H-3,4,4a), 4.39 (dd, J = 6.7, 4.1 Hz, 1H, H-2), 6.05 (ABq, J = 1.2 Hz, 2H, -OCH<sub>2</sub>O-), 6.21 (brs, 1H, NH), 6.32 (dd, J = 3, 1.2 Hz, 1H, H-1), 6.70 (s, 1H, H-10), 9.2 (s, 1H, OH).

### 2,7-Di-[(tert-butyldimethyl)-silyloxy]-narciclasine-3,4-acetonide

Diisopropylethyl amine (1.8 ml, 10.35 mmol) was added (under argon) to a heated (60°C) solution of narciclasine 3,4-acetonide (800 mg, 2.3 mmol) in dimethylformamide (8 ml) followed by tert-butyldimethylsilyl chloride (1.04 g, 6.9 mmol). The resulting reddish solution was stirred at room temperature overnight and monitored by TLC (hexane: acetone, 4:1). After completion, water (50 ml) was added and the viscous mixture was poured into ether (450 mL). The ethereal solution was washed with 10% aqueous citric acid (50 mL), water (2 x 100 mL), dried and evaporated under reduced pressure to give a gum which was

crystallized from ethanol to afford colorless flakes of disilyloxy derivative (1.2 g, 90.5%), mp. 207-9°C,  $[\alpha]_0^{30}$  +61.2°, (c, 2.5, CHCl<sub>3</sub>), IR (NaCl)  $\nu_{\text{max}}$  3250, 2952, 2930, 2857, 1676, 1480, 1381, 1362, 112, 1057, 837 cm<sup>-1</sup>, <sup>1</sup>HNMR & (CDCl<sub>3</sub>) 0.148, 0.152 (s, 6H, 2xCH<sub>3</sub>), 0.219, 0.225 (s, 6H, 2xCH<sub>3</sub>), 0.945 (s, 9H,  $C(CH_3)_3$ ), 1.335 (s, 3H,  $CH_3$ ), 1.467 (s, 3H,  $CH_3$ ), 3.969-4.019 (m, 2H, 2xCH), 4.065 (dd, J = 7.1, 5.2 Hz, 1H, CH), 4.305 (quint, J = 2.5 Hz, 1H, CH), 5.902 (brs, 1H, NH), 5.967 (d, J = 1.2 Hz, 1H, 1/20CH<sub>2</sub>0), 5.984 (d, J = 1.2 Hz, 1H, 1/20CH<sub>2</sub>0), 6.153 (brt, J = 2.3 Hz, 1H, H-1), 6.799 (s, 1H, H-10).

1,10b-(a)-and-( $\beta$ )-Epoxy-2,7-di-[(tert-butyldimethyl)silyloxy]-narciclesine-3,4-acetonide

To a stirred solution of 2,7-di-[(tert-butyldimethyl)-silyloxy]narciclasine-3,4-acetonide (240 mg, 0.42 mmol) in  $CH_2Cl_2$  (10 mL) was added 0.2M phosphate (pH 8.0) buffer (10 ml, prepared from Na2HPO, and NaH2PO,). The biphasic mixture was cooled to 0°C and m-chloroperbenzoic acid (215 mg. 1.2 mmol) was added and the mixture was stirred 20 min. at 0°C and then 4 hrs. at room temperature. The reaction was carefully monitored (TLC, hexane:acetone, 17:3) and upon completion CH<sub>2</sub>Cl<sub>2</sub> (100 mL) was added. The organic phase was separated and washed with 5% aqueous sodium thiosulfate (2 x 25 mL), water (25 mL), 5% aqueous sodium carbonate (3 x 25 mL), water (25 mL), dried  $(Na_2SO_4)$ , and evaporated under reduced pressure to produce a colorless powder. Chromatography (VLC) on neutral SiO<sub>2</sub> and elution with hexane-acetone (95:5) gave an inseparable mixture ( $\alpha$ : $\beta$ , HNMR of hydrogenolyzed product) of epoxides, (205 mg, 83.3% combined yield), mp.  $196-9^{\circ}C$ ,  $[\alpha]_0^{30} + 97.1^{\circ}$  (c, 1.05,  $CHCl_3$ ), IR (NaCl)  $\nu_{max}$  3350, 2953, 2930, 1680, 1473, 1361, 1344, 1252, 1106, 1063, 1034, 839, 777 cm<sup>-1</sup>, <sup>1</sup>HNMR & (CDCl<sub>3</sub>) 0.155 (s, 6H, SiC(CH<sub>3</sub>)<sub>2</sub>), 0.215, 0.242 (s, 3H each, SiCH<sub>3</sub>), 0.950 (s, 9H, C(CH<sub>3</sub>)<sub>3</sub>), 1.007 (s, 9H, C(CH<sub>3</sub>)<sub>3</sub>), 1.293 (s, 3H,  $1/2C(CH_3)_2$ ), 1.427 (s, 3H,  $1/2C(CH_3)_2$ ), 3.811 (s, 1H), 3.869 (d, J = 8.0Hz, 1H), 4.158-4.266 (m, 3H), 5.763 (brs, 1H, NH), 5.972 (d, 1H, J = 1.200 (m, 3H), 3.869 (m, 3H), 3.869 (m, 3H), 5.763 (brs, 1H, NH), 5.972 (d, 1H, H) 1.6 Hz, 1H,  $1/20CH_2O$ ), 6.014 (d, J = 1.6 Hz, 1H,  $1/20CH_2O$ ), 7.294 (s, 1H, H-10), EIMS (m/z) 591 (2a), 576 (10), 534 (100), 518 (4), 476 (10), 430 (10), 344 (4), 316 (6).

## (major product)

la and l $\beta$ -Hydroxy-2.7-di-[(tert-butyldimethyl)silyloxy]-10b, 4a-cis and transdihydro-narciclasine-3.4-acetonide

To a solution of epoxide mixture (see above) (100 mg, 0.17 mmol) in methanol:ethylacetate (1:3, 20 mL) was added 10% palladium supported on carbon (100 mg). The reaction mixture was evacuated and flushed with hydrogen (5x) and then hydrogenated at ambient temperature and pressure for 1 hr using a hydrogen filled balloon. The catalyst was removed by filtration and the filtrate concentrated to dryness to give a powder (97 mg), purified on PLC (SiO<sub>2</sub>, hexane:ethylacetate, 4:1) to give a mixture (1:17, <sup>1</sup>HNMR analysis) of 16, 10ba and 1a, 10bB alcohols (80 mg, 80%, found to lose the phenolic silyl group in solution), as an amorphous powder from acetone-hexane, mp 116-9°,  $[a]_0^{30}$  +7.5° (c, 0.55, CHCl<sub>3</sub>), IR (NaCl)  $\nu_{\text{max}}$  3250, 2952, 2929, 1675, 1473, 1382, 1360, 1250, 1220, 1111, 1069, 1042, 839 cm<sup>-1</sup>, <sup>1</sup>HNMR of major product (la-hydroxy isomer),  $\delta$  (CDCl<sub>3</sub>) 0.096, 0.158 (s, 3H each, Si(CH<sub>3</sub>)<sub>2</sub>), 0.231 (s, 6H,  $C(CH_3)_3$ , 1.403 (s, 3H,  $CH_3$ ), 1.531 (s, 3H,  $CH_3$ ), 2.310 (brs, 1H, OH), 2.926 (brs, 1H, H-10b), 3.745 (dd, J = 7.5, 3.3 Hz, 1H, H-2), 3.828 (brs, 1H, H-4a), 4.109 (dd, J = 5.4, 1.5 Hz, 1H, H-4), 4.196 (d, J = 4.0 Hz, 1H, H-1), 4.217 (dd, J = 7.3, 5.3 Hz, 1H, H-3), 5.115 (brs, 1H, NH), 5.930 (d, J = 1.3Hz, 1H,  $1/20CH_2O$ ), 5.993 (d, J = 1.3 Hz, 1H,  $1/20CH_2O$ ), 6.415 (s, 1H, H-1O).

#### (major product)

 $1\alpha, 2\alpha, 3\beta, 4\beta, 7$ -pentacetyl-10b, 4a-cis-dihydro-narciclasine ( $1\alpha, 10b\beta$ -isopancratistatin pentacetate) and Pancratistatin pentacetate

To a cooled  $(0^{\circ}C)$  solution of the mixture of silylether (see above, 15 mg, 0.025 mmol) in methanol (2 mL) and water (0.5 ml) was added acetic acid (0.5 ml) and trifluroacetic acid (0.5 ml). The solution was stirred at 0°C for

2 hrs and then stored in a refrigerator overnight. Solvents were removed under reduced pressure and the resulting product was dried under high vacuum over phosphorous pentoxide for 4 hrs. The product was then acetylated using pyridine (0.5 mL) and acetic anhydride (0.5 mL) at room temperature overnight followed by heating at 60°C for 1 hr. The reaction mixture was quenched with methanol and the volatile materials were evaporated through azeotropic distillation with methanol and cyclohexane. Product was found to be a (1:9) mixture of pancratistatin pentaacetate (detected only by NMR spectrum of the mixture) and  $1\alpha$ ,  $10b\beta$ -isopancratistatin pentaacetate. The products were separated on a column of silica gel by elution with CH2Cl2:CH3OH, 99:1 to give 9.0 mg of an amorphous powder from  $CH_2Cl_2$ -hexans of  $la_12a_13\beta_14\beta_17$ -pentaacetyl-10b, 4a-cis-dihydro-narciclasine, mp.165-9°,  $[\alpha]_0^{30}$  +135 (c, 0.2, CHCl<sub>3</sub>), IR (NaCl)  $\nu_{\text{max}}$  3341, 1778, 1751, 1677, 1481, 1371, 1248, 1226, 1192, 1084, 1035 cm-1, 1HNMR & (CDCl<sub>3</sub>) assignment based on 1H, 1H-COSY spectra, 1.893, 1.979, 2.027, 2.166, 2.351 (each s, 3H each, 5 x OCOCH<sub>3</sub>), 3.305 (c, J = 3.8 Hz, 1H, H-10b), 3.933 (r, J = 2.5 Hz, 1H, H-4a), 5.405 (dd, J = 10.7, 3.2 Hz, 1H, H-2), 5.460 (brs, 1H, NH), 5.470 (dd, J = 10.7, 2.3 Hz, 1H, H-3), 5.488 (brs, 1H, H-4), 5.532 (c, J - 3.4 Hz, 1H, H-1), 6.068 (d, J - 1.2 Hz, 1H,  $1/20CH_2O$ ), 6.085 (d, J = 1.2 Hz, 1H,  $1/20CH_2O$ ), 6.626 (s, 1H, H-1O), the chemical shift for NH shifted downfield at  $\delta$  5.590 in dilute solutions (ca. 1.5 mg/0.5 mL). Cis relationship of the protons at H-4a, H-10b and H-1 established by NOE measurement. Thus strong NOE's were observed between H-10b, H-1, H-2, H-10, H-4a, and H-4a also gave NOE enhancement to NH). The NOE's also establishes proof for the chair conformation of ring C.

(major product)

#### la-Isopancratistatin

Palladium/carbon (10%, 80 mg) was added to the epoxide mixture described above (80 mg, 0.14 mmol) in anhydrous THF (45 mL) and the hydrogenolysis was performed as described in the previous experiment for 8 hrs. The filtrate obtained after removal of the catalyst was concentrated to dryness to give a 2:1 mixture of trans:cis dihydro product, by HNMR analysis. The products were separated on PLC (hexane-acetone (17:3) to give trans dihydro product (42 mg, 52.3%) and cis dihydro product (20 mg, 24.8%), identical (HNMR and TLC) with the cis product obtained in the previous hydrogenolysis reaction. Slightly impure trans dihydro product was obtained as an amorphous powder from acetone-hexane; HNMR 6 (CDCl<sub>3</sub>) of major product: 0.104, 0.138, 0.190, 0.199 (each s, 3H each, 2 x Si(CH<sub>3</sub>)<sub>2</sub>), 0.902, 0.971 (each s, 9H each, 2 x C(CH<sub>3</sub>)<sub>3</sub>), 1.312, 1.433 (each s, 3H each, C(CH<sub>3</sub>)<sub>2</sub>), 2.814 (dd, J = 14.3, 7.7 Hz, 1H, H-10b), 3.085 (d, J = 3.7 Hz, 1H, 0H), 3.457 (dd, J = 14.1, 8.0 Hz, 1H, H-4a), 3.866 (dd, J = 7.2, 5.0 Hz, 1H, H-2), 4.055 (m, 1H, H-1), 4.207 (t, J = 8.5

Hz, 2H, H-3,4), 5.696 (s, 1H, NH), 5.910 (d, J = 1.2 Hz, 1H,  $1/20\text{CH}_20$ ), 5.950 (d, J = 1.2 Hz, 1H,  $1/20\text{CH}_20$ ), 6.998 (s, 1H, H-10).

To a ccoled (0°C) solution of trans product (25 mg 0.042 mmol) in THF:CH<sub>3</sub>OH:H<sub>2</sub>O (1.5:2:1, 4.5 mL) was added acetic acid (0.5 mL) and trifluoroacetic acid (1.0 mL) and stirred at the same temperature for 1 hr. After storing overnight in the refrigerator, the reaction was not complete and required heating to 40°C for 8 hrs. Solvents were removed under reduced pressure and the product was purified by flash chromatography on silica gel. The product, la-isopancratistatin (11.1 mg, 81%), eluted with a 9:1 mixture of CH<sub>2</sub>Cl<sub>2</sub>:CH<sub>3</sub>OH and was obtained as an amorphous powder, mp. 325-7, IR (KBr)  $\nu_{\rm max}$  3500-3300, 1679, 1470, 1337, 1285, 1210, 1141, 1089, 1064, 802, 725 cm<sup>-1</sup>, <sup>1</sup>HNMR & (DMSO-d<sub>8</sub>+D<sub>2</sub>O) 2.80 (dd, 10.9, 10.9 Hz, 1H, H-10b), 3.31 (dd, J = 13.2, 10.5 Hz, 1H, H-4a), 3.76 (m, 2H), 3.81 (t, J = 3.6 Hz, 1H), 3.84 (brs, 1H), 5.97, 6.00 (only two AB lines visible, 2H, OCH<sub>2</sub>O), 7.27 (s, 1H, H-10).

1a,2a,3\$,4\$,7-Pentascetyl isopancratistatin

la-Isopancratisatin (2.7 mg) was treated with acetic anhydride (0.2 mL) in pyridine (0.2 ml) at 50°C for 2 hrs. The mixture, quenched with methanol, was reduced to dryness under a nitrogen stream. Product was chromatographed on a column of silica gel and eluced with  $CH_2Cl_2:CH_3OH$  (49:1) to give an amorphous powder of la-isopancratistatin pentaacetate (3.4 mg, 76.5%), mp. 146-8, IR (NaCl)  $\nu_{max}$  3350, 2930, 2850, 1755, 1676, 1482, 1370, 1248, 1226, 1176, 1084, 1059, 1033 cm<sup>-1</sup>, <sup>1</sup>HNMR & (CDCl<sub>3</sub>) 2.06 (s, 3H, COCH<sub>3</sub>), 2.09 (s, 3H, COCH<sub>3</sub>), 2.12 (s, 3H, COCH<sub>3</sub>), 2.18 (s, 3H, COCH<sub>3</sub>), 2.36 (s, 3H, COCH<sub>3</sub>), 3.44 (t, J = 11.9 Hz, 1H, H-10b), 3.84 (t, J = 11.0 Hz, 1H, H-4a), 5.25 (dd, J = 10.8,

3.0 Hz, 1H, H-4), 5.42 (dd, J = 11.4, 3.3 Hz, 1H, H-1), 5.44 (t, J = 3.3 Hz, 1H, H-1), 5.53 (t, J = 3.8 Hz, 1H, H-2), 6.06 (d, J = 1.2 Hz, 1H,  $1/20CH_2O$ ), 6.07 (d, J = 1.2 Hz, 1H,  $1/20CH_2O$ ), 6.54 (s, 1H, H-10), spectrum was assigned on the basis of 2D-COSY, and the stereochemistry by NOEDS data.

la-Hydroxy-2-[(tert-butyldimethyl)silyloxy]-10b,  $4a-\underline{cis}$  and  $\underline{trans}$ -isopancratistatin-3,4-acetonide and 1a-hydroxy-2-[(tert-butyldimethyl)silyloxy]- $\Delta(10b,4a)$ -isopancratistatin-3,4-acetonide

The silyloxy epoxide mixture (150 mg) was dissolved in THF (10 mL) and hydrogenolyzed using hydrogen-10% Pd/C (50 mg) as described. Chromatography on a silica gel column and elution with hexane-acetone (7:3) gave 7-desilylated products (interestingly desilation was occurring during the hydrogenolysis reaction as crystallized epoxide was free of benzoic acid by NMR) trans:cis: $\Delta(10b,4a)$  in the ratio of (5:3:5). The 10b,4a trans product (50 mg, 38%), crystallized from methanol as shining flakes, mp. 274-5; IR (NaCl)  $\nu_{\rm max}$  3530, 3360, 2952, 2939, 1678, 1466, 1373, 1361, 1345, 1260, 1230, 1085, 1071 cm<sup>-1</sup>; <sup>1</sup>H NMR & (CDCl<sub>3</sub>) 0.15 (s, 3H, SiCH<sub>3</sub>), 0.18 (s, 3H, SiCH<sub>3</sub>), 0.94 (s, 9H, SiC(CH<sub>3</sub>), 1.37 (s, 3H, CH<sub>3</sub>), 1.48 (s, 3H, CH<sub>3</sub>), 2.90 (dd, J = 14.3, 6.9 Hz, 1H, H-10b), 3.13 (d, J = 2.7 Hz, 1H, 0H), 3.57 (dd, J = 14.5, 7.9 Hz, 1H, H-4a), 3.92 (dd, J = 6.8, 5.2 Hz, 1H, H-2), 4.13 (ddd, J = 7.4, 4.6, 2.2 Hz, 1H, H-1), 4.25 (t, J = 6.7 Hz, 1H, H-4), 4.29 (t, J = 6.7 Hz, 1H, H-3), 6.03 (ABq, J = 3.0 Hz, 2H, OCH<sub>2</sub>0), 6.04 (brs, 1H, NH), 6.93 (s, 1H, H-10), 12.48 (s, 1H, ArOH), (assignment was made on the basis of a 2D-COSY analysis and stereochemical assignment was accomplished by NOEDS measurement).

Continued elution of the column with the same solvent gave a mixture of cis and 10b,4a (A) product. The cis product could not be separated

but crystallization of the mixture from methanol yielded pure 10b, 4a ( $\Delta$ ) olefinic product (55 mg, 41.4%), recrystallized from methanol as flakes, mp. 266-8; IR (NaCl)  $\nu_{max}$  3535 (brs), 2989, 2959, 2931, 2897, 2857, 1677, 1625, 1485, 1422, 1373, 1253, 1215, 1117, 1086, 1036 cm<sup>-1</sup>; <sup>1</sup>H NMR 6 (CDCl<sub>3</sub>) 0.17 (s, 3H, SiCH<sub>3</sub>), 0.21 (s, 3H, SiCH<sub>3</sub>), 0.97 (s, 9H, SiC(CH<sub>3</sub>), 1.49 (s, 3H, CH<sub>3</sub>), 1.50 (s, 3H, CH<sub>3</sub>), 2.90 (s, 1H, OH), 3.86 (dd, J = 7.8, 3.5 Hz, 1H, H-2), 4.50 (t, J = 7.5 Hz, 1H, H-3), 4.76 (d, J = 3.4 Hz, 1H, H-1), 5.11 (d, J = 7.0 Hz, 1H, H-4), 6.10 (d, J = 1.6 Hz, 1H, 1/20CH<sub>2</sub>0), 6.11 (d, J = 1.6 Hz, 1H, 1/20CH<sub>2</sub>0), 6.83 (s, 1H, H-10), 9.85 (s, 1H, NH), 12.65 (s, 1H, OH), Assignment is based on 2D-COSY spectrum and stereochemistry was determined by NOEDS measurement.

Hydrogenolysis of the silyloxy epoxide mixture on a scale better than the one reported here in different solvents (ethyl acetate, mixture of ethyl acetate and methanol) produced similar products. Hydrogenolysis in methanol mostly produced the  $\Delta$  (10b, 4a) product.

## 2,7-Diacetoxy-narciclasine-3,4-acetonide

Acetonide was prepared from narciclasine (1 g) as described before and all the solvents were evaporated under reduced pressure to give a crude product which was acetylated with acetic anhydride (3 mL) - pyridina (3 mL) at  $60^{\circ}$ C for 6 hrs. Solvents were evaporated under reduced pressure after addition of methanol and then chromatographed on a silica gel column and eluted with hexane-ethyl acetate-methylene chloride (3:1:2) to give pure diacetate (1.2 g, 85.48) as an amorphous powder from acetone-hexane, mp. 130-33 °C; IR (NaCl)  $\nu_{max}$  3350, 1775, 1745, 1671, 1482, 1373, 1233, 1210, 1177, 1081, 1031 cm<sup>-1</sup>; <sup>3</sup>H NMR  $\delta$  (CDCl<sub>3</sub>) 1.39 (s, 3H, CH<sub>3</sub>), 1.52 (s, 3H, CH<sub>3</sub>), 2.21 (s, 3H, COCH<sub>3</sub>), 2.39 (s, 3H, COCH<sub>3</sub>), 4.12 (dd, J = 7.8, 7.8 Hz, 1H, H-3), 4.16 (brs, 1H, H-4a), 4.31 (dd, J = 7.5, 5.8 Hz, 1H, H-4), 5.39 (dd, J = 4.9, 2.3 Hz, 1H, H-2), 6.02 (brs, 1H, NH), 6.09 (brs, 2H, OCH<sub>2</sub>O), 6.12 (t, J = 3.2 Hz, 1H, H-1), 6.98 (s, 1H, H-1O).

# 1,10b-(a)-Epoxy-2,7-diacetoxy-narciclasine-3,4-acetonide

To a solution of narciclasine acetonide diacetate (1.0 g, 2.32 mmol) in  $CH_2Cl_2$  (60 mL) was added 0.2 M soadium phosphate buffer (pH 8, 60 mL) followed by m-chloroperbenzoic acid (1.4 g, 3.5 molar equivalent). The reaction mixture was stirred at room temperature overnight and then  $CH_2Cl_2$  (500 mL) was added and the organic layer was separated, washed with 5% solution of sodium thiosulfate (3 x 200 mL), 5% solution of sodium carbonate (3 x 200 mL), water (2 x 100 mL), dried ( $Na_2SO_4$ ) and evaporated to give almost pure epoxide as a powder, crystallized from acetone-hexane (700 mg, 67.5%) as amorphous granules, mp. 231-232 °C; IR (NaCl)  $\nu_{max}$  3370, 1792, 1749, 1682, 1500, 1365, 1345, 1209, 1175, 1083, 1032 cm<sup>-1</sup>; H NMR & ( $CDCl_3$ ) 1.31 (s, 3H,  $CH_3$ ), 1.43 (s, 3H,  $CH_3$ ), 2.20 (s, 3H,  $COCH_3$ ), 2.36 (s, 3H,  $COCH_3$ ), 4.00 (d, J = 6 Hz, 1H, H-4a), 4.03 (s, 1H, H-1), 4.27 (apparent t, J = 8.1 Hz, H-1, H-3), 4.38 (dd, J = 7.9, 6.3 Hz, 1H, H-4), 5.33 (d, J = 6.1 Hz, 1H, H-2), 5.82 (brs, 1H, NH), 6.07 (ABq, J = 1.2 Hz,  $OCH_2O$ ), 6.43 (s, 1H, H-10).

Hydrogenation of 7-deoxynarciclasine. A solution of 7-deoxynarciclasine (1.02 g, 3.4 mmol) in methanol-ethanol (400 ml, 1:1) was degassed with nitrogen, platinum oxide (57 mg) was carefully added and the resulting mixture was hydrogenated at ambient temperature and pressure for 24 hrs. The reaction mixture was filtered through Celite and concentrated in vacuo to afford crude iso-7-deoxynarciclasine (150 mg, dark brown solid) which crystallized from pyridine-hexane as a powder (100 mg, 9.8% yield). Identity with earlier sample of the compound was confirmed by mp and nmr. The mother liquor contained cis and trans-dihydro-7-deoxynarciclasine.

cis-dihydro-7-deoxynarciclasine

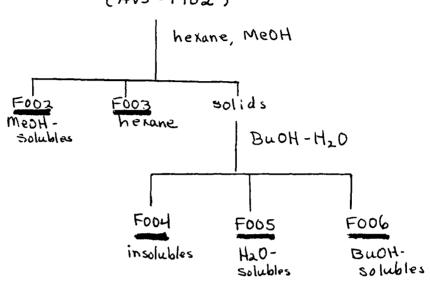
cis and trans-dihydronarciclasine triacetate. The crystallization residue from <u>iso-7-deoxynarciclasine</u> (see above) was concentrated to dryness and treated with acetic anhydride (7 ml) and pyridine (10 ml) at  $60^{\circ}$  for 6 hrs. Methanol was added and the resulting solution was concentrated to dryness. The mixture was flash chromatographed over silica gel using  $\text{CH}_2\text{Cl}_2$ :MeOH (99.4-0.6) twice to furnish <u>trans</u>-dihydro-7-deoxynarciclasine (112 mg, 7.6% yield) and <u>cis-</u>dihydro-7-deoxynarciclasine (752 mg, 51.2% yield). Identity with earlier sample of the compound was confirmed by mp and nmr.

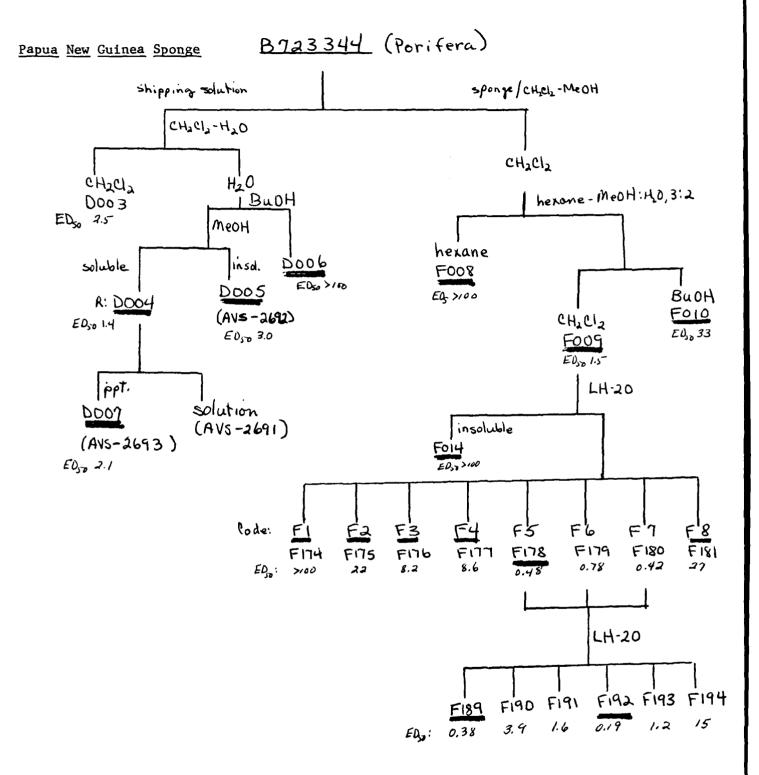
Trans-dihydro-7-deoxynarciclasine. To a solution of the triacetate (112 mg) in methanol (20 ml) was added a saturated solution of barium hydroxide (6 ml). After heating for 15 min. at  $100^{\circ}$ C, the mixture was cooled, saturated with solid  $CO_2$ , stirred at room temperature overnight and filtered. The filtrate was evaporated to dryness and the product was crystallized from methanol to give trans-dihydro-7-deoxynarciclasine (51 mg, 65% yield). Identity with earlier sample of the compound was confirmed by mp and nmr.

cis-dihydro-7-deoxymarciclasine. To a solution of the triacetate (750 mg) in methanol (80 ml) was added potassium carbonate (300 mg). The mixture was stirred at room temperature for 2 hrs, then filtered through a column of Sephadex LH-20. Elution with  $\text{CH}_2\text{Cl}_2$ :MeOH (3:2) removed the product from the column. Crystallization from acetone-MeOH afforded cis-dihydro-7-deoxynarciclasine as crystals (419 mg, 80% yield). Identity with earlier sample of the compound was confirmed by mp and nmr.

Philippines Sponge

B721160, Haliclona sp. (Porifera)
aqueous extract
(AVS-1902)





Grant No. DAMD17-89-Z-9021

#### CONCLUSIONS

The discovery that pancratistatin will cure USAMRIID's <u>in vivo</u> Japanese Encephalitis has opened the way to a new generation of antiviral drugs. Doubtlessly, current efforts at uncovering new naturally occurring antiviral drugs will lead to analogous excellent progress.

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